

LETTER

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# Immune biomarker-based enrichment in sepsis trials

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Dear Editor

We read with great interest the study by Anderson et al. recently published in *Critical Care* [1]. The study assessed whether soluble mediators (IL-8, sTNFR1, and Ang-2) could be used as biomarkers to “enrich” subject populations with higher mortality risk in subsequent clinical trials. The authors addressed “immunocompetence” of patients using clinical parameters (APACHE-II score and/or presence of ARDS). They found that both IL-8 and sTNFR1 (but not Ang-2) were suitable for identification of patients with higher mortality risk and concluded that IL-8 and sTNFR1 can be used as “prognostic enrichment factors” in future clinical sepsis studies.

Biomarker-based prognostic enrichment appears important to select sample populations with a greater likelihood of having improved clinical outcomes following a given therapeutic intervention. Although sTNFR1 and IL8 levels may be associated with higher mortality in certain sepsis patients, however, selection of the correct “enrichment markers” should be performed cautiously and based on a solid underlying biological rationale. This may, for example, be of particular importance in the field of clinical sepsis trials testing immunomodulatory interventions where assessment of the pleiotropic cytokine IL-8 would likely introduce considerable bias and should thus not be used to stratify respective patient populations. Failure of an adequate peri-interventional characterization may at least partly explain the failure of a number of previous sepsis trials testing immunological interventions (e.g., corticosteroids, strategies testing anti-TNF or anti-LPS). In the study by Anderson et al., “immunocompetency” in sepsis patients was defined using clinical criteria. However, it seems that immunocompetency, i.e., (functional) immune phenotype, cannot be assessed by predominantly clinical parameters and

should be based on comprehensive functional immune markers (e.g., mHLA-DR expression [2–4]) in order to identify individuals who would benefit most from a given intervention.

We are well aware that the focus of the article was to address the important question of whether biomarker-based enrichment would lead to better stratification of future trial cohorts.

While we appreciate the insights provided by Anderson et al., we believe that it will be crucial (and challenging) to continue the quest for the “correct” enrichment markers to succeed in the design of novel therapeutic interventions, which may require more extensive reverse translational research and personalized treatment approaches [5]. In the light of failure of a large number of previous clinical sepsis trials, it seems apparent that biomarker enrichment using appropriate mediators is needed and may open several avenues towards more personalized treatment approaches in sepsis.

## Abbreviations

IL-8: Interleukin 8; sTNFR1: Soluble tumor necrosis factor receptor-1; Ang-2: Angiopoietin-2; APACHE : Acute Physiology, Age, Chronic Health Evaluation; ARDS: Acute Respiratory Distress Syndrome; TNF: Tumor necrosis factor; LPS: Lipopolysaccharide

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TS and JCS drafted and finalized the manuscript. CM and SvG helped to draft the manuscript and revised the manuscript for important intellectual content. All authors read and approved the final version of the manuscript.

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### References

1. Anderson BJ, Calfee CS, Liu KD, Reilly JP, Kangelaris KN, Shashaty MGS, Lazaar AL, Bayliffe AI, Gallop RJ, Miano TA, et al. Plasma sTNFR1 and IL8 for prognostic enrichment in sepsis trials: a prospective cohort study. *Crit Care*. 2019;23(1):400.
2. Pfortmueller CA, Meisel C, Fux M, Scheffold JC. Assessment of immune organ dysfunction in critical illness: utility of innate immune response markers. *Intensive Care Med Exp*. 2017;5(1):49.
3. Monneret G, Lepape A, Voirin N, Bohe J, Venet F, Debard AL, Thizy H, Bienvenu J, Gueyffier F, Vanhems P. Persisting low monocyte human leukocyte antigen-DR expression predicts mortality in septic shock. *Intensive Care Med*. 2006;32(8):1175–83.
4. Scheffold JC. Measurement of monocytic HLA-DR (mHLA-DR) expression in patients with severe sepsis and septic shock: assessment of immune organ failure. *Intensive Care Med*. 2010;36(11):1810–2.
5. von Gunten S. The future of pharmacology: towards more personalized pharmacotherapy and reverse translational research. *Pharmacology*. 2019; 105(1–2):1–2.

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